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**Biosupplies Australia Pty Ltd**

P.O. Box 187 La Trobe University, Bundoora  
Victoria 3083 AUSTRALIA

**Email** [info@biosupplies.com.au](mailto:info@biosupplies.com.au)

**Web** [www.biosupplies.com.au](http://www.biosupplies.com.au)

ABN 62 006 209 034



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## Histochemical Reagents

### Aniline blue fluorochrome for callose

An analytical probe for (1→3)-β-D-glucans (callose). Highly specific for (1→3)-β-glucans [Evans et al. (1984) *Carbohydrate polymers* **4**: 215-230; Stone et al. (1984) *Protoplasma* **122**: 191-195]. May be used for the quantitative determination of callose [Kauss (1989) *Plant Physiol.* **81**: 171-176] and in fluorescence assays for (1→3)-β-glucan synthase products.

### Yariv reagents for arabinogalactan-proteins (AGPs)

Specific probes for the detection of AGPs in tissue sections [Anderson et al. (1977) *Aust. J. Plant Physiol.* **4**: 143-158]. May be used to detect and quantify AGPs in tissue extracts [van Holst & Clarke (1985) *Anal Biochem.* **148**: 446-450] and to detect AGPs in crossed-electrophoretic separations. [van Holst & Clarke (1986) *Plant Physiol.* **80**: 786-798].

### Gum arabic: an arabinogalactan-protein (AGP)

Gum Arabic AGP is used as a reference standard in many methods involving detection, quantification and analytical analyses of AGP's from plant tissues.

## Immunohistochemical Reagents

### Monoclonal antibodies (Murine)

#### To (1→3)-β-D-glucan

- No cross-reactivity with (1→4)-β-D-glucans or (1→3,1→4)-β-D-glucans [Meikle et al. (1991) *Planta* **188**: 1-8].

#### To (1→3,1→4)-β-D-glucan

- No cross-reactivity with (1→3)-β-D-glucans or (1→4)-β-D-glucans [Meikle et al. (1994) *The Plant Journal* **5**: 1-9].

#### To (1→4)-β-D-mannan and galacto-(1→4)-β-D-mannan

- [Pettolino et al. (2001). *Planta* **214**: 235-242]

#### To arabinogalactan-proteins (under licence from University of Leeds, UK)

- Arabinogalactan-proteins, terminal glucuronosyl residues (LM2) [Smallwood et al. (1996). *Planta* **198**: 452-459.]
- Arabinogalactan-proteins, β-GlcA-(1-3)-α-GalA-(1-2)-Rha epitope (JIM13) [Knox et al. (1991). *Plant Journal* **1**: 317-326.]

#### To heteroxylans (under licence from University of Leeds, UK)

- (1-4)-β-D-xylan (LM11) [McCartney et al. (2005). *J. Histochemistry & Cytochemistry* **53**(4): 543-546.]
- Heteroxylan, glucuronoxylan (LM28) [Cornuault et al. (2015). *Planta* **242**(6): 1321-1334.]

#### To xyloglucans (LM25) (under licence from University of Leeds, UK)

- [Pedersen et al. (2012). *J. Biological Chemistry* **287**(47): 39429-39438.]

#### To pectic polysaccharides (under licence from University of Leeds, UK)

- Pectic (1-5)-α-L-arabinan (LM6) [Willats et al. (1998). *Carbohydrate Research* **308**(1-2): 149-152.]
- Branched pectic galactan (LM26) [Torode et al. (2018). *Plant Physiology* **176**(2): 1547-1558.]
- Pectic (1,4)-β-D-galactan (LM5) [Jones et al. (1997). *Plant Physiology* **113**(4): 1405-1412.]
- Pectic unesterified homogalacturonan (LM19) [Verherbruggen et al. (2009). *Carbohydrate Research* **344**(14): 1858-1862.]
- Pectic methylesterified homogalacturonan (LM20) [Verherbruggen et al. (2009). *Carbohydrate Research* **344**(14): 1858-1862.]
- Pectic methylesterified homogalacturonan (JIM7) [Knox et al. (1990). *Planta* **181**(4): 512-521.]

#### To extensins (under licence from University of Leeds, UK)

- Extensin (JIM20) [Smallwood et al. (1994). *The Plant Journal* **5**(2): 237-246.]
- Extensin (LM1) [Smallwood et al. (1995). *Planta* **196**(3): 510-522]

Each of these antibodies can be used with second stage, gold- or fluorochrome-labelled rabbit, anti-mouse antibody for immunohistochemical studies.

## Enzymes

### (1→3,1→4)-β-D-Glucan Hydrolase from *Bacillus subtilis* (EC 3.2.1.73)

Specifically hydrolyses β-D-glucans containing both (1→3) and (1→4)-β-D-glucosidic linkages in linear sequences. Does not hydrolyse (1→4)-β-D-glucans or (1→3)-β-D-glucans. [Anderson & Stone (1975) *FEBS Letters* **52**: 202-207].

## Substrates

### Pachyman [(1→3)-β-D-glucan] ex *Poria cocos*

Useful as a positive control in fluorescence microscopy studies on callose using the aniline blue fluorochrome or (1→3)-β-D-glucan specific monoclonal antibody.



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