

# Human FGF-21 (Luc) HEK293 Reporter Cell Data Sheet

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# Human FGF-21 (Luc) HEK293 Reporter Cell Data Sheet

## Human FGF-21 (Luc) HEK293 Reporter Cell

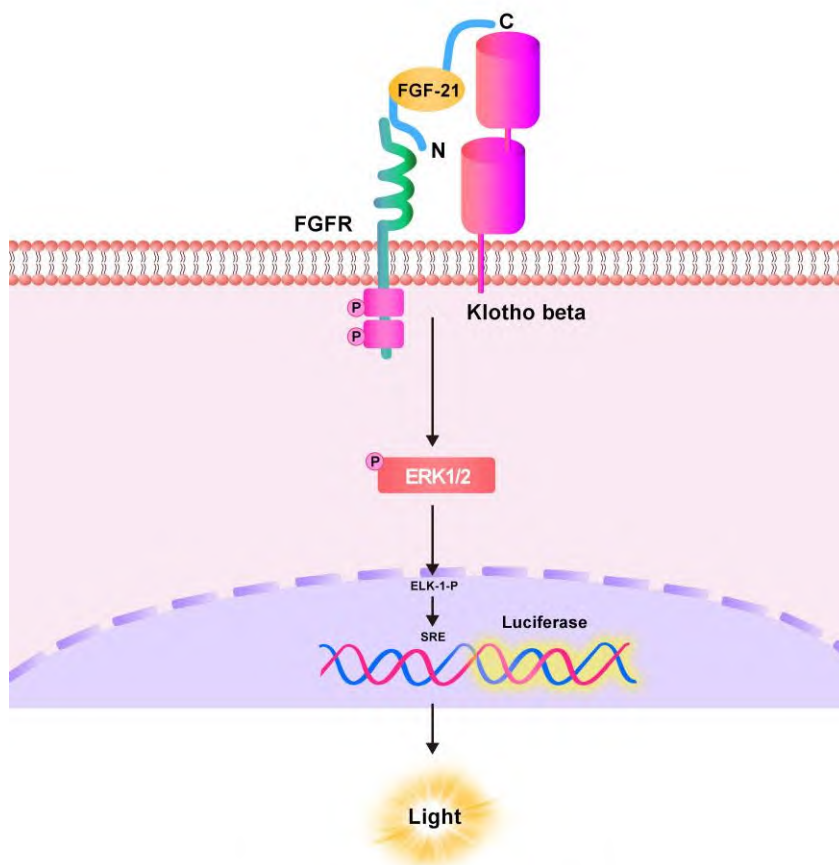
Catalog No.	Size
CHEK-ATF163	2 × (1 vial contains ~5×10 <sup>6</sup> cells)

### • Description

The Human FGF-21 (Luc) HEK293 Reporter Cell was engineered to not only express SRE signaling response element, but also express the receptor human Klotho beta (Gene ID: 152831). When stimulated with human FGF-21 protein, receptor-mediated signaling can drive SRE-mediated luminescence.

### • Application

- Bioactivity detection of human FGF-21 fusion protein.



Human FGF-21 (Luc) HEK293 Reporter Cell

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## • Cell Line Profile

Cell line	Human FGF-21 (Luc) HEK293 Reporter Cell
Host Cell	HEK293
Property	Adherent
Complete Growth Medium	DMEM + 10% FBS
Selection Marker	Puromycin (2 µg/mL) + Hygromycin B (20 µg/mL)
Incubation	37°C with 5% CO <sub>2</sub>
Doubling Time	22-24 hours
Transduction Technique	Lentivirus

## • Materials Required for Cell Culture

- DMEM Medium (BasalMedia, Cat. No. L120KJ)

**Note:** If you are unable to obtain the specified DMEM medium (BasalMedia, Cat. No. L120KJ) in China, you may use an alternative DMEM medium (Gibco, Cat. No. 11965-092) or another suitable medium for culturing.

- Fetal bovine serum (CellMax, Cat. No. SA211.02)
- Puromycin (InvivoGen, Cat. No. ant-pr-5b)
- Hygromycin B (Invitrogen, Cat. No. 10687010)
- 0.25% Trypsin-EDTA (1X), Phenol Red (Gibco, Cat. No. 25200-056)
- Penicillin-Streptomycin (Gibco, Cat. No. 15140-122)
- Phosphate Buffered Saline (1X) (HyClone, Cat. No. SH30256.01)
- Complete Growth Medium: DMEM + 10% FBS, 1% P/S
- Culture Medium: DMEM + 10% FBS, Puromycin (2 µg/mL), Hygromycin B (20 µg/mL), 1% P/S
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, Cat. No. 430641)
- Cryogenic storage vials (SARSTEDT, Cat. No. 72.379.007)
- Thermostat water bath
- Centrifuge (Cence, Model: L550)
- Cell counter (MONWEI, Model: SmartCell200A Plus)
- CO<sub>2</sub> Incubator (Thermo, Model: 3111)
- Biological Safety Cabinet (Thermo, Model: 1389)

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## • *Recovery*

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 70% ethanol. All the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 4.0 mL complete growth medium and spin at approximately 1000 rpm for 5 minutes.
4. Resuspend cell pellet with 5 mL **complete growth medium** and transfer the cell suspension into T-75 flask containing 10-15 mL of pre-warmed complete growth medium.
5. Incubate at 37°C with 5% CO<sub>2</sub> incubator until the cells are ready to be split.

## • *Subculture*

1. Remove and discard culture medium.
2. Wash the cells once with sterile PBS.
3. Add 2 mL of 0.25% trypsin to cell culture flask. Place the flask at 37°C for 2-3 minutes, until 90% of the cells have detached.
4. Add 6.0 to 8.0 mL of **culture medium** and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new culture vessel.
6. Incubate at 37°C with 5% CO<sub>2</sub> incubator.

**Subcultivation Ratio:** A subcultivation ratio of 1:6 to 1:10 is recommended.

**Medium Renewal:** Every 2 to 3 days.

**Note:** After recovery for 1-2 generations with the complete growth medium not containing the selection marker, if the cell state is well, changing to the culture medium containing the selection marker.

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## • *Cryopreservation*

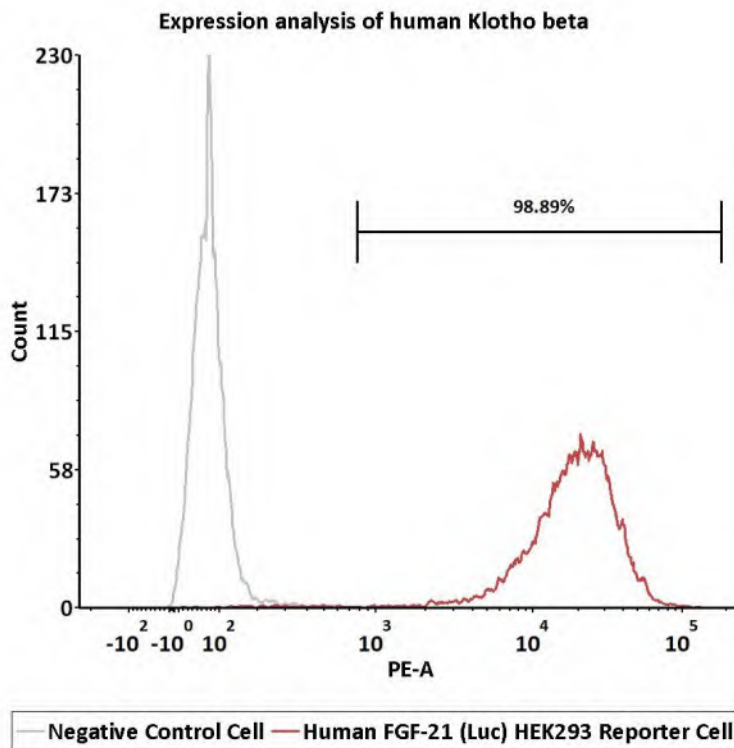
1. Remove and discard spent medium.
2. Detach cells from the cell culture flasks with 0.25% trypsin.
3. Centrifuge at 1000 rpm for 5 min at RT to pellet cells.
4. Resuspend the cell pellets with complete growth medium and count viable cells.
5. Centrifuge at 1000 rpm for 5 min at RT and resuspend cells in freezing medium to a concentration of  $5 \times 10^6$  to  $1 \times 10^7$  cells/mL.
6. Aliquot into cryogenic storage vials. Place vials in a programmable cooler or an insulated box placed in a  $-80^{\circ}\text{C}$  freezer overnight, then transferring to liquid nitrogen storage.

## • *Storage*

- **Product format:** Frozen
- **Storage conditions:** Liquid nitrogen immediately upon receipt

# Human FGF-21 (Luc) HEK293 Reporter Cell Data Sheet

## • Receptor Assay

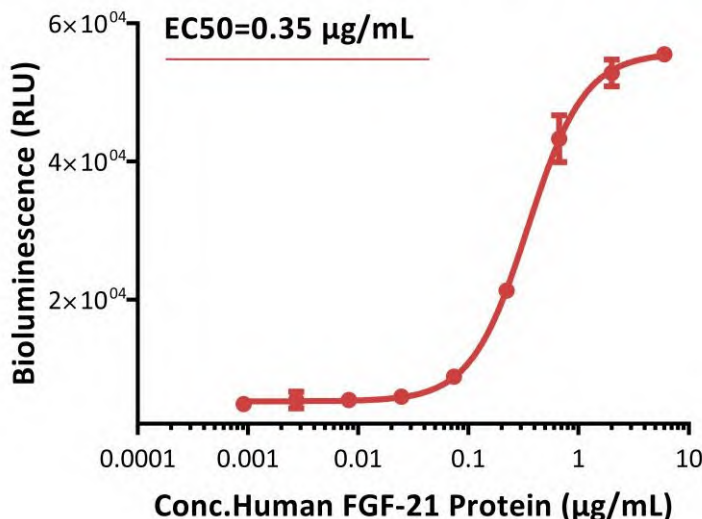


**Fig1. Expression analysis of human Klotho beta on Human FGF-21 (Luc) HEK293 Reporter Cell by FACS.** Cell surface staining was performed on Human FGF-21 (Luc) HEK293 Reporter Cell or negative control cell using anti-human Klotho beta antibody followed by staining with PE anti-human IgG antibody.

# Human FGF-21 (Luc) HEK293 Reporter Cell Data Sheet

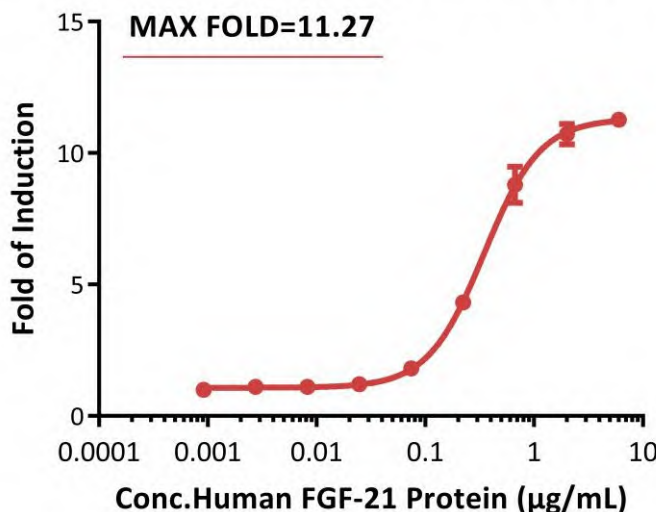
## • Signaling Bioassay

### Human FGF-21 protein Stimulation (RLU)



**Fig2. Response to human FGF-21 protein (RLU).** The Human FGF-21 (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human FGF-21 protein (Cat. No. FG1-H5243). The EC50 was approximately 0.35 µg/mL.

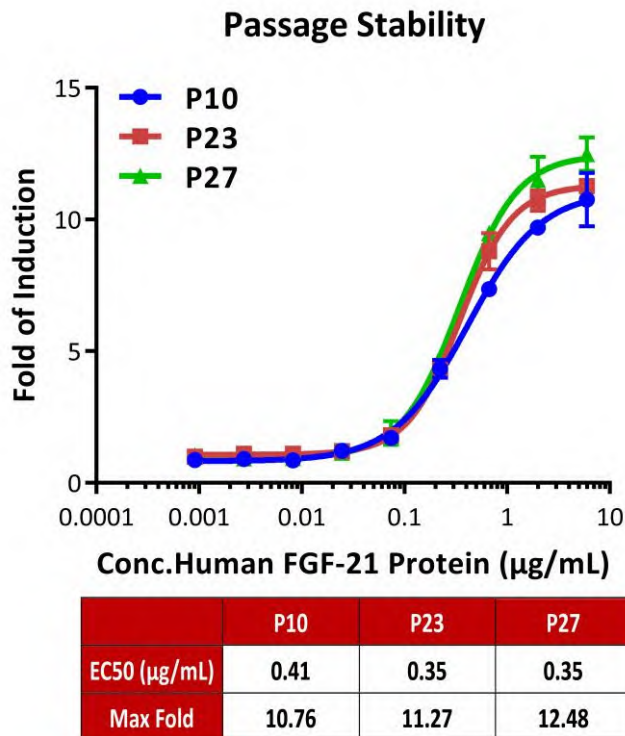
### Human FGF-21 protein Stimulation (FOLD)



**Fig3. Response to human FGF-21 protein (FOLD).** The Human FGF-21 (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human FGF-21 protein (Cat. No. FG1-H5243). The max induction fold was approximately 11.27.

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## • Passage Stability



**Fig4. Passage stability analysis by Signaling Bioassay.** The continuously growing Human FGF-21 (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human FGF-21 protein (Cat. No. FG1-H5243). Human FGF-21 protein stimulated response demonstrates passage stabilization (fold induction and EC50) across passage 10-27.

# Human FGF-21 (Luc) HEK293 Reporter Cell Data Sheet

## • *Related Products*

<b><u>Products</u></b>	<b><u>Cat. No.</u></b>
Human FGF-21 Protein, His Tag	FG1-H5243
Human THRB (Luc) HEK293 Reporter Cell	CHEK-ATF181
Human THRA (Luc) HEK293 Reporter Cell	CHEK-ATF180
Human GLP-1R (Luc) HEK293 Reporter Cell	CHEK-ATF096
Human GCGR (Luc) HEK293 Reporter Cell	CHEK-ATF103
Human GIPR (Luc) HEK293 Reporter Cell	CHEK-ATF104
HEK293/Human GLP-1R Stable Cell Line (High Expression)	CHEK-ATP160
HEK293/Human GLP-1R Stable Cell Line (Medium Expression)	CHEK-ATP161
HEK293/Human GLP-1R Stable Cell Line (Low Expression)	CHEK-ATP162
HEK293/Human GLP-1R&GIPR Stable Cell Line	CHEK-ATP205
HEK293/Human GIPR Stable Cell Line (High Expression)	CHEK-ATP206
HEK293/Human GIPR Stable Cell Line (Medium Expression)	CHEK-ATP207
HEK293/Human GIPR Stable Cell Line (Low Expression)	CHEK-ATP208
HEK293/Human GCGR Stable Cell Line (High Expression)	CHEK-ATP209
HEK293/Human GCGR Stable Cell Line (Medium Expression)	CHEK-ATP210
HEK293/Human GCGR Stable Cell Line (Low Expression)	CHEK-ATP211
Human Activin RII (Luc) HEK293 Reporter Cell	CHEK-ATF164
HEK293/Human ASGR1&ASGR2 Stable Cell Line	CHEK-ATP172
HEK293/Human GPR75 Stable Cell Line	CHEK-ATP174
HEK293/Human ASGR1 Stable Cell Line	CHEK-ATP080