His MultiTrap TALON

Product Information

Cat#No#	Hi-400C

Product Overview

His MultiTrap TALON is a ready-to-use 96-well filter plate for high reproducibility in expression screening and small-scale parallel purification of histidine-tagged proteins.

Description

His MultiTrap TALON multiwell plates are designed for high-throughput screening of histidine-tagged proteins from clarified cell lysates. His MultiTrap TALON prepacked plates are manufactured with high reproducibility with regards to the well and resins volumes in order to enable consistent results. The average purity of His MultiTrap TALON is > 85% with yields up to 1 mg/well. Leakage of cobalt ions from TALON Superflow resin is negligible. His MultiTrap TALON is used manually with vacuum or centrifugation, or in a liquid handling station.

Characteristic

Up to 1 mg of pure histidine-tagged protein per well.
Prefilled with 50 µL TALON Superflow cobalt-IMAC (immobilized metal affinity chromatography) resin for reproducible and high throughput parallel screening, offering a different selectivity compared to nickel-charged resins.
Large sample volumes can be applied all at once.

Particle Size

60 to160 µm

Dynamic binding capacity

Up to 1 mg histidine-tagged protein/ well

Chemical compatibility

Stable in all commonly used buffers, reducing agents, denaturants, and detergents.

pH working range

2–14

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CIP stability
3–12
Storage
4 to 8°C, 20% Ethanol
Binding buffer
The recommended binding buffer condition is at neutral to slightly alkaline pH (pH 7 to 8) in the presence of 0.3 M to 0.5 M NaCl. Sodium phosphate buffers are often used.
Elution buffer
Imidazole is used for elution of histidine-tagged proteins.
Purification procedures
1. Remove storage solution: Remove the bottom seal. Gently shake the 96-well filter plate while holding it upside down, to remove any medium stuck on the top seal. Place the plate in upright position. Remove the top seal from the plate while holding it against the bench surface. Position the plate on top of a collection plate. 2. Prewash: : Add 500 µl deionized water/well. Centrifuge for 2 minutes at 500 × g. 3. Equilibrate: Add 500 µl binding buffer/well and mix briefly, to equilibrate the medium. Centrifuge for 2 minutes at 500 × g. Repeat once. 4. Load sample: Apply sample to the wells (maximum 600 µl/well). Incubate for 3 minutes. (Increase the incubation time if the yield is too low). Remove the flowthrough by centrifuging for 4 minutes at 100 × g (or until all wells are empty). 5 Wash:

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Add 500 µl washing buffer/well to wash out unbound sample.
Centrifuge for 2 minutes at 500 × g.
Repeat once (or until all unbound sample are removed).
6. Elute:
Add 200 µl of elution buffer/well and mix for 1 minute.
Change collection plate and centrifuge the plates for 2 minutes at 500 × g and collect the fractions.
Repeat twice (or until all target protein has been eluted).
If required, change collection plate between each elution.

Pack size
4 × 96-well filter plates
Column volume
800 µl

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