

4. Run NADPH assay in supernatants reaction:

4.1 Add 50 μL of NADPH reaction mixture (from Step 2) into each well of NADPH standard, blank control, and test samples (see Step 3.3) to make the total NADPH assay volume of 100 μL /well

Note 1: For a 384-well plate, add 25 μL of sample and 25 μL of NADPH reaction mixture into each well.

Note 2: Prepare cells or tissue samples as desired. Lysis Buffer (Component D) can be used for lysing the cells for convenience (See appendix for details).

4.2 Incubate the reaction at room temperature for 15 minutes to 2 hours, protected from light.

4.3 Monitor the absorbance increase with an absorbance plate reader at 460 nm.

Data Analysis

The absorbance in blank wells (with the PBS buffer only) is used as a control, and is subtracted from the values for those wells with the NADPH reactions. A NADPH standard curve is shown in Figure 1.

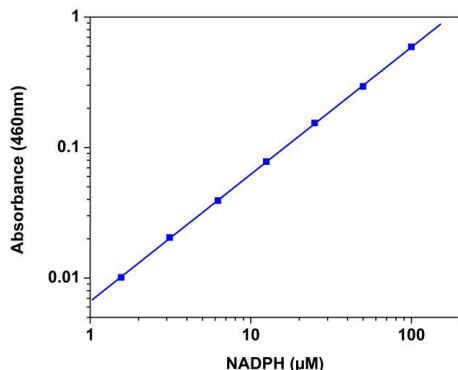


Figure 1. NADPH dose response was measured with Amplitude™ Colorimetric NADPH Assay Kit in a 96-well white/clear bottom plate using a SpectraMax microplate reader (Molecular devices). As low as 3 μM of NADPH can be detected with 30min incubation (n=3) with absorbance measurement at 460nm.

Appendix: Test Sample Preparations Using Component D (Lysis Buffer)

- Plant Cell Samples:** Homogenize leave with the lysis buffer at 200 mg/mL, and centrifuge at 2500 rpm for 5-10 minutes, use the supernatant for tests.
- Bacterial Cell Samples:** Collect bacterial cells by centrifugation ((10,000 g, 0°C, 15 min). Use about 100 to 10 million cells/mL lysis buffer, keep the treated solution at room temperature for 15 minutes. Centrifuge at 2500 rpm for 5 minutes, and use the supernatant for tests.
- Mammalian Cell Samples:** Remove medium from plate wells, use about 100 μL lysis buffer per 1-5 million cells (or 50-100 μL /well in a 96-well cell culture plate), and keep the treated solution at room temperature for 15 minutes. Use the cell lysate directly or centrifuge it at 1500 rpm for 5 minutes, use the supernatant for tests.
- Tissue Samples:** Weigh ~20 mg tissue, wash with cold PBS. Homogenize with 400 μL of lysis buffer in a micro-centrifuge tube. Centrifuge at 2500 rpm for 5-10 minutes, use the supernatant for the assay.

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